





INTERNATIONAL JOURNAL OF ENGINEERING SCIENCES & RESEARCH TECHNOLOGY

THE DECOMPOSER MICROORGANISMS IN THE ENVIRONMENT AND THEIR SUCCESSION OF SUBSTRATES

Raj Singh*, Anju Rani, Permod Kumar, Gyanika Shukla, Amit Kumar

* Department of Botany, Faculty of Science, Swami Vivekanand Subharti University, Meerut, UP, India

DOI: 10.5281/zenodo.58637

ABSTRACT

Plant litter decomposition is not a purely chemical or physical process, it is basically a biological one resulting from the diverse activities of microorganisms, protozoa and various other soil organisms like insects and worms. The bacteria and fungi play a very significant role in plant litter decomposition and humus formation. The fungi which colonized decaying substrates could use only simple substrates such as sugars and they referred as sugar fungi. Garrett suggested that the root infecting fungi open the way for a sequence of saprophytic sugar fungi, cellulose decomposers and finally lignin decomposers. The initial rates of losses of differrent components could be ranked as sugar > hemicelluloses> cellulose> lignin. During this succession, the substrates were found to become progressively depleted. This biochemically based succession was believed to be reflected in a taxonomic way as the Phycomycetes to be considered as sugar fungi, the Ascomycetes and Deuteromycetes as cellulolytic and finally Basidiomycetes as lignin decomposers.

KEYWORDS: Plant litter, decomposition, microorganisms, cellulose, hemicelluloses, lignin.

INTRODUCTION

Decompsition and Humification: A central process in the life cycle of all organisms is the decomposition of their dead remains. Plants are no exception to it. Truly speaking, a significant proportion of the net primary production in the biosphere, especially in the terrestrial ecosystems, ends up as accumulations of dead remains of plants at soil surface or below it. These dead remains get decomposed; and the nutrients locked up in the litter as well as a number of new substances formed in the process of decomposition, pass down the soil profile. The significance of this process was recognized long ago.

One may find references to "humus" and its importance in increasing soil fertility in the literature of 18th centuary¹. Remann² established that humus formation is not a purely chemical or physical process; rather, it is basically a biological one resulting from the diverse activities of microorganisms, protozoa and various other soil organisms like insects and worms. Mitscherlich³ was the first to attribute the decomposition of organic material to bacteria. Since that time, and till the beginning of the twentieth century, the bacteria were considered to be the chief agents responsible for the decomposition of organic matter in the soil. The role of fungi in the formation of the brown substance of the humus was recognized by Scherpe⁴. Dascewska⁵ demonstrated experimentally that the Hyphomycetes play a significant role in the decomposition of cellulose and in humification. Singh and Charaya⁶ isolated and identified about 40 fungi belonging to Hypomycetes from decomposing wheat crop resedues.

Plant litter decomposition and microorganisms: The fungi play a "predominant" role in the decomposition of plant litter, is now a well-established fact. It is now realised that the plant litter is decomposed by a sequence of events involving physical processes like leaching and mechanical breakdown; as well as through biological processes like microbial degradation which involve several exo-enzymes⁷.

In aerobic environments, the saprobic fungi which can secrete cellulases and other exo-enzymes directly into the environment constitute a major group amongst the decomposer community^{8,9,10}. In the aquatic environment also, the decomposition of plant litter is predominantly carried out by the fungi—the fungal biomass being much greater than



[Singh* *et al.*, 5(7): July, 2016] ICTM Value: 3.00

ISSN: 2277-9655 Impact Factor: 4.116

the bacterial biomass. Using the rates of incorporation of 14C into ergosterol and that of 3H leucine into protein as indices for estimating fungal and bacterial biomass respectively, Suberkropp and Weyers¹¹ found higher turnover rates for fungi (11-26 times) as compared to bacteria, in the decomposing leaves in streams thereby further implicating a major role for the fungi in litter decomposition. Dilly¹² studied the culturable bacterial and fungal communities during the decomposition of leaf litter in a black alder forest. Their studies also revealed that fungal communities play a predominant role in litter break down. Earlier, Mamilov¹³ had suggested an improved substrate-induced respiration method for differential determination of fungal and bacterial biomass in the soil upon the decomposition of plant remains, suggesting the use of high dose of antibiotics. Extensive use of this method may throw further light on this issue.

In fact, the soil is the home of an indigenous microflora or is merely a resting place for fungal spores floating in the atmosphere. Different workers indicated that many fungi grow and reproduce in the soil. Since then, numerous studies on different aspects of soil microorganisms have been made and it is now almost impossible to bring even the complete list of all such studies under a single cover. There are also numerous articles, reviews and books concerning soil microflora these include^{14,15,16,17,18,19,6,20}.

Succession of decomposers: Earlier studies on soil fungi were mainly concerned with the compilation of floristic lists. However, gradually the emphasis shifted on studying the ecology of soil mycobiota. The synecological studies led to a survey of substrate relationships of these fungi. Garrett²¹ suggested that it is the root-infecting fungi which open the way for a sequence of saprophytic 'sugar fungi', cellulose-decomposers and finally lignin-decomposers in the invaded tissue. Later on, Garrett²² proposed the following very generalised scheme for the succession of fungi on a corpus of plant tissue lying within or upon the soil.

Senescent tissue	Dead tissue		
Stage I a	Stage I	Stage II	Stage III
Weak parasites	Primary saprophytic sugar	Cellulose decomposers and	Lignin decomposers
	fungi, living on sugar and	associated secondary saprophytic	and associated fungi
	carbon compounds similar	sugar fungi, sharing products of	
	than cellulose	cellulose decomposition	

Table 1: Succession of fungi

In this scheme, Garrett recognized a 'secondary group' of 'sugar fungi' which do not live on simple carbon sources initially present in the fresh plant tissue, but are dependent upon the hydrolytic products of cellulose-decomposing fungi; and, therefore, appear late in succession in association of these. One of the features that led to the development of substrate-group hypothesis was the observation that the initial rates of losses of different components could be clearly ranked: Sugar > Hemicellulose > Cellulose > Lignin.

During this succession, the substrates were found to become progressively depleted, beginning with sugars and the simpler carbon compounds, continuing with cellulose; and finally with lignin. Similar succession was observe by Charaya and Singh²³, Talbot²⁴ and Singh²⁵ during the decomposition of wheat crop residues. This biochemically based succession was believed to be reflected in a taxonomic one in so far as the Phycomycetes (no more recognised as a taxonomic group) came to be considered as 'sugar fungi' *par excellence*; the Ascomycetes and Fungi Imperfect as cellulolytic; the decomposition of lignin was believed to be primarily the work of the higher Basidiomycetes²⁶. The succession of fungus fruiting bodies on herbivore dung experimentally analysed by Harper and Webster²⁷. The observations by a number of workers like Saito²⁸ on beech litter and (Sundaram²⁹) on a variety of substrates. Sinha and Dayal³⁰ on teak lend support to the Garrett's scheme. The succession observed by Chang and Hudson³¹; Singh and Charaya⁶ on wheat straw compost also followed this scheme in broadest outline, but the Phycomycetous phase was found to be of very short duration.

On lignicolous substrates, a rather different type of succession was observed by a number of workers like Mangenot³² on trunks of deciduous trees; Meredith³³ on pine stumps. In general, the substrates were initially found to be colonized by Basidiomycetes and Fungi Imperfecti, the Phycomycetes appearing later. Thus, the general trend of succession was reversed with an initial phase of cellulose and lignin- decomposing fungi. Studies by Caldwell³⁴ on beech litter also



[Singh* et al., 5(7): July, 2016] ICTM Value: 3.00

ISSN: 2277-9655 Impact Factor: 4.116

demonstrated that Phycomycetes did not play any significant role in the decomposition of woody substrates. Frankland³⁵ followed the succession of fungi on decaying petioles of bracken (*Pteridium aquilinum*). The following trend proposed by her had two interesting features : (i) the occurrence of potential lignin-decomposers (Basidiomycetes) and Sphaeropsidales early in the succession and; (ii) the main build up of Mucorales in the end : Hyphomycetes + (Sphaeropsidales ----> Basidiomycetes ---> Phycomycetes). The occurrence of Sphaeropsidales in the early phase of succession was also reported by Hering³⁶ on leaves of oak, hazel and ash; Macauley³⁷ on *Eucalyptus* spp. in different environments.

On other substrates, the pattern of succession appears to be one or several waves of Ascomycetes and Fungi Imperfecti following each other. In fact, Chesters³⁸ embarked upon the idea that the aerial parts of "plant shoot systems are worked over" by weak parasitic and saprobic species, and they arrive at the soil surface much depleted of their nutritional possibilities. Kendrick^{39,40} found initial colonization of pine litter by weak parasites. The studies of the saprobic mycobiota of Saccharum officinarum by Hudson⁴¹ revealed a primary fungal biota consisting of a number of weak parasites and saprobes followed by Periconiella and some imperfect fungi; and finally by numerous imperfect fungi and Ascomycetes. More or less similar pattern was observed by Hogg⁴² on leaves of *Fagus sylvatica*; Singh and Charaya⁶ on wheat crop resedue, Hudson⁴³ suggested that for many types of leaf and herbaceous litters, some general correspondence of pattern could be discerned in the observed changes. He proposed the following scheme for the succession of fungi on most plant debris except the lignicolous ones:

Table 2: Succession of Jungi				
Living tissue	Senescent	Dead		
Parasites:	Common primary	Secondary saprophytes:		
Ascomycetes and fungi imperfecti may	saprophytes: Ascomycetes and	1. Ascomycetes and fungi		
be host specific or host restricted	fungi imperfecti	imperfecti		
	Restricted primary	2. Basidiomycetes		
	saprophytes: Ascomycetes and	3. Soil inhabitants (Mucorales,		
	fungi imperfecti	Penicillia etc.		

Ruscoe⁴⁴ found that the leaves of *Nothofagus truncatus* were already heavily colonized by a variety of parasitic and saprobic fungi when they reached the forest floor. Tubaki and Yokoyama⁴⁵) studied the succession on the leaves of Castanopsis cuspidata and Quercus phyllyraeoides. While confirming the Hudson's scheme, they observed four distinct stages in succession-(i) A group of early colonizers comprising the transient fungi, present on the leaf surface only as detachable propagules, (ii) fungi growing and sporulating actively through out the decay period; and (iii) a group found early in the decay process, but disappearing later; and (iv) a group found at late stages.

Studies by Shearwood and Carroll⁴⁶ on Pseudotsuga meziesti; Aneja and Mehrotra⁴⁷ on Desmostachya litter. Singh and Charaya⁶ on wheat crop residues, more or less corroborate the pattern of succession proposed by Hudson⁴³. On the basis of a scrutiny of the investigations of different workers, Earlier, Charaya⁴⁸ studied the mycobiota colonising wheat and paddy straw aboveground, at soil surface (control and waterlogged conditions), and beneath soil surface (control and water-logged conditions). Fungal succession on wheat and paddy roots buried underground were also studied (in control as well as in water-logged conditions). The pattern of succession on both types of roots under both conditions was as suggested by Hudson. On wheat straw also, the succession basically followed the pattern as suggested by Hudson, but allowing for the incorporation of 'secondary sugar fungi' of Garrett²² at intervals. On paddy straw, however, initial colonisation by the Phycomycetes was observed (though alongwith Ascomycetes and Hyphomycetes); the Phycomycetes disappeared sooner or later. In case of the paddy straw decomposing at soil surface under water-logged conditions, even the Ascomycetes disappeared in the end; the fungal succession in these cases, thus, followed the pattern suggested by Garrett²². The study, thus, suggested that the pattern of colonization of a substrate by the mycoflora is largely regulated by the nature of the resource itself⁴⁹. The mycoflora naturally occurring in paddy straw was studied by Singh⁵⁰ without any attempt to study the pattern of successive colonization. Selvam⁵¹ studied the microbial colonization of decomposing paddy straw using nylon bag technique. Singh and Charaya⁶ also studied the microbial colonization of different components of wheat crop resedues viz., wheat internodes, leaves, chaff and mixed straw.

http://www.ijesrt.com

THOMSON REUTERS ENDNOTE [Singh* et al., 5(7): July, 2016] ICTM Value: 3.00

ISSN: 2277-9655 Impact Factor: 4.116

CONCLUSION

The urbanization and industrialization responsible for hudge amount of solid wastes including plant litters and house hold organic wastes, causes environmental problems. Microbes are able to degrade such type of lignocellulosic wastes convert them into products of social welfare such as biogas, enzymes, organic acids etc. This study analysis the biochemical and taxonomic succession of fungi of decomposing plant litters. It is helpful to decide the pattern of colonization of substrates by decomposers and to develop a microbial technology for lignocellulosic waste recycling for safe and clean environment.

ACKNOWLEDGEMENT

I my express my profound sense of gratitude and indebtedness to my esteemed supervisor Professor Dr. M.U. Charaya who with high tenacity, subverted all the snages in the progress of this work and has throughout been a constant source of motivation, imagination and information. I also express my heart-felt gratitude to Professor V.K. Bhatnager, Pro Vice Chancelor, Swami Vivekananda Subharti University, Meerut and Professor Rita Bakshi, Dean Faculty of Science of this University for their ever encouraging suggestions, besides providing me all the necessary facilities to complete this significant work. I express my profound thanks to my friends and colleagues for their co-operation and assistance in this work.

REFERENCES

- [1] Achard, F. Chemische Untersuchung des Torf's. Grell's Chem. Ann. 2, 391 (1786).
- [2] Remann, E. Bodenkunde, 2nd Ed., Berlin (1905).
- [3] Mitscherlich, E. Monatsber d. K. Press, Akad. d., Wiss. Zu. Berlin, 102 (Quoted by Macbeth and Scales in U. S. Dept. Agr. Buv. Plant Ind. Bull., 1913, pp. 266 (1850).
- [4] Scherpe, R. Uber den Einflus des Schwelkohlenst offs auf die stickstoofumset-zungsvorgange in Boden. Arb. K. Gsndhtsamt. *Biol. Abt.* **7**, 353-428 (1909).
- [5] Dascewska, W. Etude sur la desaggregation de la cellulose dans la terre de bruyeue et la toube. *Univ. Geneva Inst. Bot.* S 8, fas. 8, 239-316 (1913).
- [6] Singh, R and Charaya M.U. Fungal colonization of decomposing above ground residues of wheat crop. *Bulletin of Pure and Applied Sciences* 22B(1), 55-59(2003).
- [7] Sinsabaugh, R. L., Benfield, E. F. & Linkins, A. E. Cellulase activity associated with the decomposition of leaf litter in a woodland steram. *Oikos* **36**, 184-190(1981).
- [8] Kshatriya, S., Sharma, G.D. & Mishra, R.R. Enzyme activities related to litter decomposition in forests of different age and altitude in North East India. *Soil Biol. Biochem.* **24**, 265-270 (1992).
- [9] Singh R., Charaya M.U., Shukla L., Shukla G., Kumar A., and A. Rani. "Lignocellulolytic Potentials of Aspergillus terreus for Management of Wheat Crop Residues". Journal of Academia and Industrial Research, 3(9), 453-455 (2015a).
- [10] Rani, A., Girdharwal, V., Singh, R., Kumar, A. & Shukla, G. "Production of Laccase enzyme by white rot fungi Coriolus versicolor". Journal of Environmental and Applied Bioresearch. **03**(04), 204-206 (2015).
- [11] Suberkropp, K. & Weyers, H. Application of fungal and bacterial production methodologies to decomposing leaves in streams. *Applied and Environmental Microbiology* **62**, 1610-1615(1996).
- [12] Dilly, O., Bartsch, S., Rosenbrock, P., Buscot, F. & Munch, V Shifts in physiological capabilities of the microbiota during the decomposition of leaf litter in a black alder (*Alnus glutinosa* Gaertn. L.) forest. *Soil Biology and Biochemistry* 33, 921-930 (2001).
- [13] Mamilov, A. Sh., Byzov, A. B., Stepanov, A. L. & Zvyagintsev, D. G. Differential determination of fungal and bacterial biomass in the soil, upon the decomposition of plant remains. *Pochvovedenie* 12, 1457-1462 (2000).
- [14] Garrett, S. D. Pathogenic Root Infecting Fungi. Cambridge University Press, London (1970).
- [15] Barron, G.L. *The Genera of Hyphomycetes from Soil*. Robert E. Krieger Publishing Co., Huntington, N. Y (1972).
- [16] Domsch, K.H., Gams, W. & Anderson, T. Compendium of Soil Fungi. Vol. I & II. Academic Press, London, N.Y., Sydney (1980).
- [17] Paul, E. A. & Clark, F. E. Soil Microbiology and Biochemistry. 2nd Edn. (1996) Academic Press, California.

http://www.ijesrt.com



[Singh* et al., 5(7): July, 2016]

ICTM Value: 3.00

ISSN: 2277-9655

Impact Factor: 4.116

- [18] Snow, K. M. Soil Microbiology. In "Encyclopedia of Microbiology" (Eds. Joshua Lederberg), 4, 321-335(2000). Academic Press, New York.
- [19] Mukerji, K. G. Soil microbes. In "*Techniques in Mycorrhizal Studies*" (Eds. Mukerji, K. G., Manoharachary, C. and B. P. Chamola). : 7-13 (2002). Kluwer Academic Publishers. Dordrecht, The Netherlands.
- [20] Espersch" J., Zimmermann C., D"umig A., G. Welzl, F. Buegger, M. Elmer, J. C. Munch, & Schloter, M. Dynamics of microbial communities during decomposition of litter from pioneering plants in initial soil ecosystems. *Biogeosciences* 10, 5115–5124 (2013).
- [21] Garrett, S. D. Biology of Root-infecting Fungi. Cambridge University Press, Cambridge (1956).
- [22] Garrett, S. D. Soil Fungi and Soil Fertility. Pergamon Press, Oxford (1963).
- [23] Charaya M.U. & Singh R. Biochemical changes in wheat crop residues during their decomposition in nature. Journal of Acta Ciencia Indica **31**(1), 39-46 (2005).
- [24] Talbot, J.M., Yelle, D.J., Nowick, J., Treseder, K.K. Litter decay rates are determined by lignin chemistry. *Biogeochemistry*, **108**, 279-295(2012).
- [25] Singh,R., Rani,A., Kumar, A. & Girdharwal, V. "Biochemical changes during in vitro decomposition of wheat residue of Trichoderma lignorum (Tode) Harz. International Journal of Advanced Information Science and Technology. 4(8), 29-3 (2015d).
- [26] Garrett, S. D. Ecological groups of soil fungi: a survey of substrate relationships. *New Phytol.* **50**, 149-166 (1951).
- [27] Harper, J. E. & Webster, J. An experimental analysis of the coprophilous fungal succession. *Trans. Br. mycol. Soc.* **47**, 511-530 (1964).
- [28] Saito, T. Microbiological decomposition of beech litter. Ecol. Rev. (Sendai) 14, 141-147 (1956).
- [29] Sundaram, B.M. Saprophytic colonization of the substrates buried in soils by fungi. *Ind. J. Microbiol.* 17, 38-40 (1977).
- [30] Sinha, A. & Dayal, R. Fungal decomposition of teak leaf litter. Indian Phytopath. 36, 54-57 (1983).
- [31] Chang, Y. & Hudson, H.J. The fungi of wheat straw compost- I. Ecological studies. *Trans. Br. mycol. Soc.* 50: 649-666 (1967).
- [32] Mangenot, F. Recherches methodiques sur les champignons de certains biois en decompositions. *Rev. gen. Bot.* **59**, 45 (1952).
- [33] Meredith, D. S. Further observations of fungi inhabiting pine stumps. Ann. Bot. 24, 63 78 (1960).
- [34] Caldwell, R. Observations on the fungal flora of decomposing beech litter in soil. *Trans. Br. mycol. Soc.* **46**, 249-261 (1963).
- [35] Frankland, J.C. Succession of fungi on decaying petioles of Pteridium quilinum. J. Ecol. 54, 41-63 (1966).
- [36] Hering, T.F. Fungal decomposition of oak leaf litter. Trans Brit. mycol. Soc. 50: 267-273 (1967).
- [37] Macauley, B.J. Biodegradation of Eucalyptus leaf litter fungi. (Abs.) *International Symposium on Microbial Ecology*, Dunedin, Newzealand. B.33 (1977).
- [38] Chesters, C.G.C. Concerning fungi inhabiting the soil. Trans. Br. mycol. Soc. 32, 197 (1949).
- [39] Kendrick, W. B. Microfungi in pine litter. *Nature* (London) **181**, 432 (1958a).
- [40] Kendrick, W. B. Microfungi of pine litter. Ph.D. Thesis, University of Liverpool (1958b).
- [41] Hudson, H.J. Succession of microfungi on ageing leaves of Saccharum officinarum. Trans. Br. mycol. Soc. 45, 395-423 (1962).
- [42] Hogg, B.M. & Hudson, H. J. Microfungi on leaves of *Fagus sylvatica*. I. The microfungal succession. *Trans. Br. mycol. Soc.* 49, 185-192 (1966).
- [43] Hudson, H.J. The ecology of fungi on plant remains above the soil. New Phytol. 67, 837-874 (1968).
- [44] Ruscoe, Q.W. Mycoflora of living and dead leaves of Nothofagus truncata. Trans. Br. Mycol. Soc. 56, 463 474 (1971).
- [45] Tubaki, K. & Yokoyama, T. Successive fungal flora on sterilized leaves in the litter of forests. III. *IFO Res. Comm.* 6, 27-49(1973 b).
- [46] Shearwood, M. & Carroll, G. Fungal succession on needles and young twigs of old growth *Douglas fir. Mycologia* **65**, 499-506 (1974).
- [47] Aneja, K.R. & R.S. Mehrotra Studies on microorganisms decomposing aboveground parts of "the grass" (*Desmostachya bipinnata*). *Proc. Nat. Acad. Sci. India* **50B**, 12-20 (1980).



[Singh* et al., 5(7): July, 2016]

ICTM Value: 3.00

ISSN: 2277-9655 Impact Factor: 4.116

- [48] Charaya, M.U. *Taxonomical, ecological and physiological studies on the mycoflora decomposing wheat and paddy crop residues.* Ph.D. Thesis. Dept. of Botany, M.M. Postgraduate College, Modinagar (Meerut University, Meerut), India (1985).
- [49] Charaya, M.U. & Mehrotra, R. S. Microbial colonisation of decomposing plant litter. In "New Trends in Microbial Ecology" (Eds. Rai, B. and M. S. Dkhar): 76-89 (1998). Dept. of Botany, NEHU, Shillong and ISCNR, Dept. of Botany, BHU, Varanasi, India.
- [50] Singh, K.P., Gupta, S.R. & Edward J.C. Fungi associated with different types of decomposing organic matter with special reference to cellulose decomposition. *The Allahabad Farmer* **50** : 259-269 (1979).
- [51] Selvam, K., Palaniswamy, M., Guruswamy, R. & Swaminathan, K. Biodegradation of forest and agrowastes in a social forestry ecosystem. In "*Microbes for better living*" (Eds. Sankaran, R. and K. S. Manja): 313-321(1995). Conference Secretariat, Micon International-94, Defence Food Research Laboratory, Mysore-570011. India.